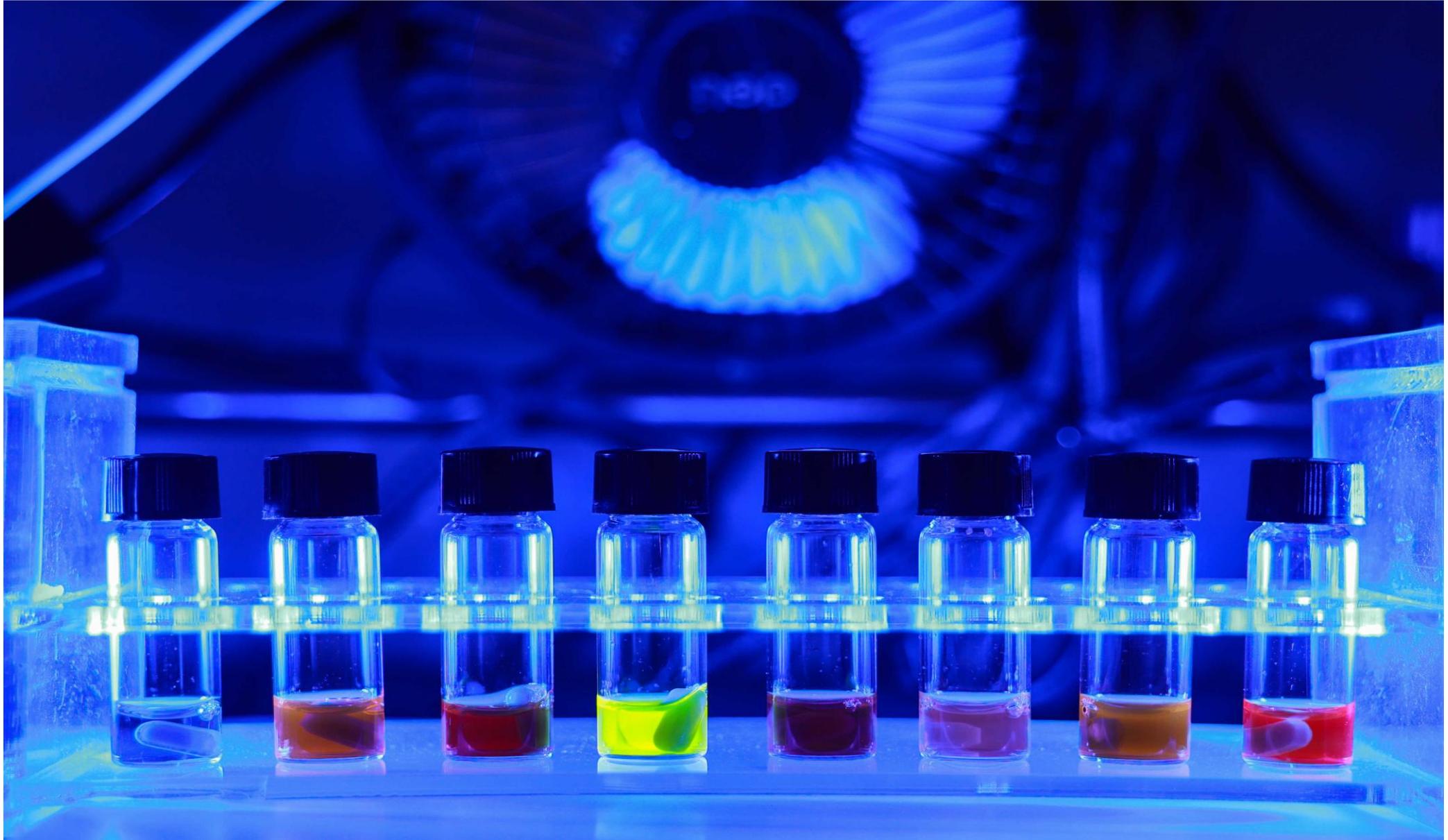


Literature Report



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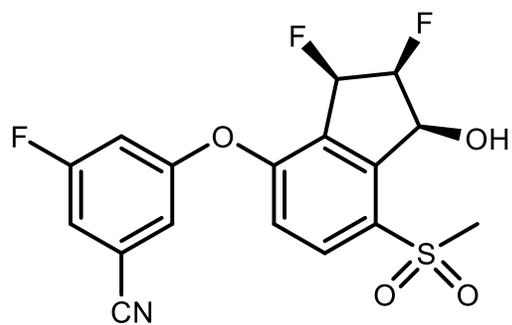
2

Photoenzymatic radical hydroalkylation

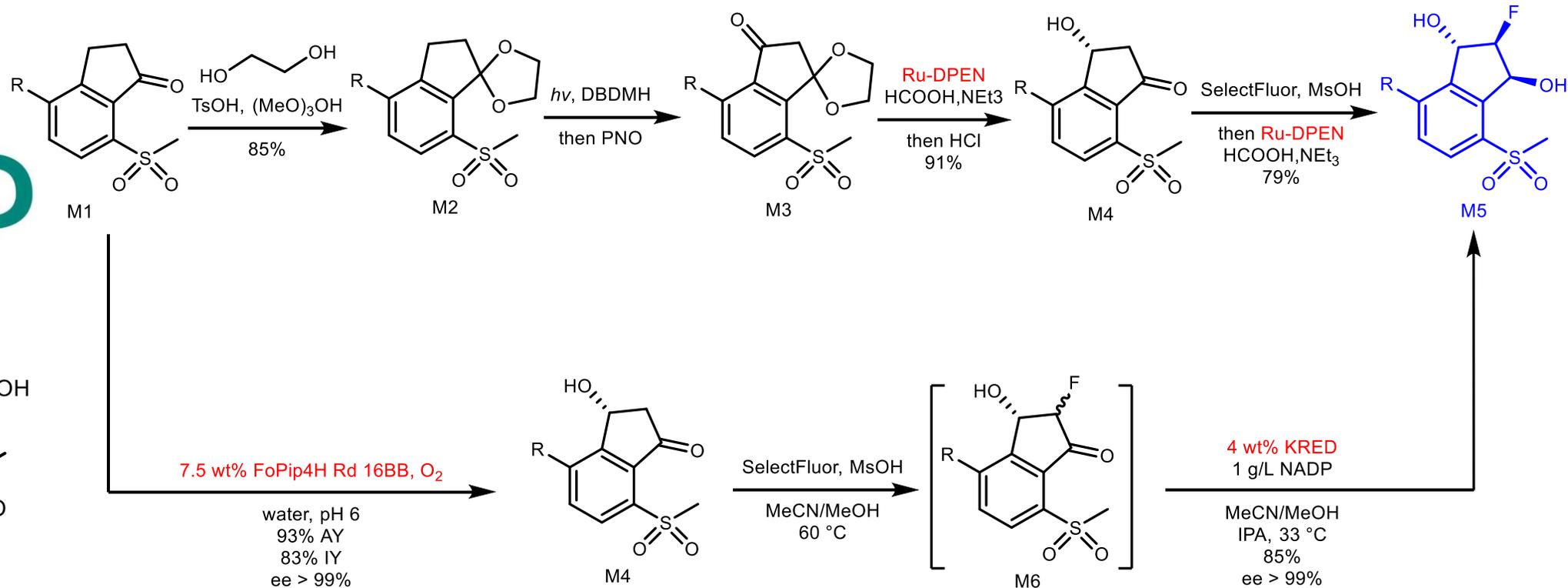
3

Conclusion

The application of enzymatic catalysis in Pharmaceutical Research

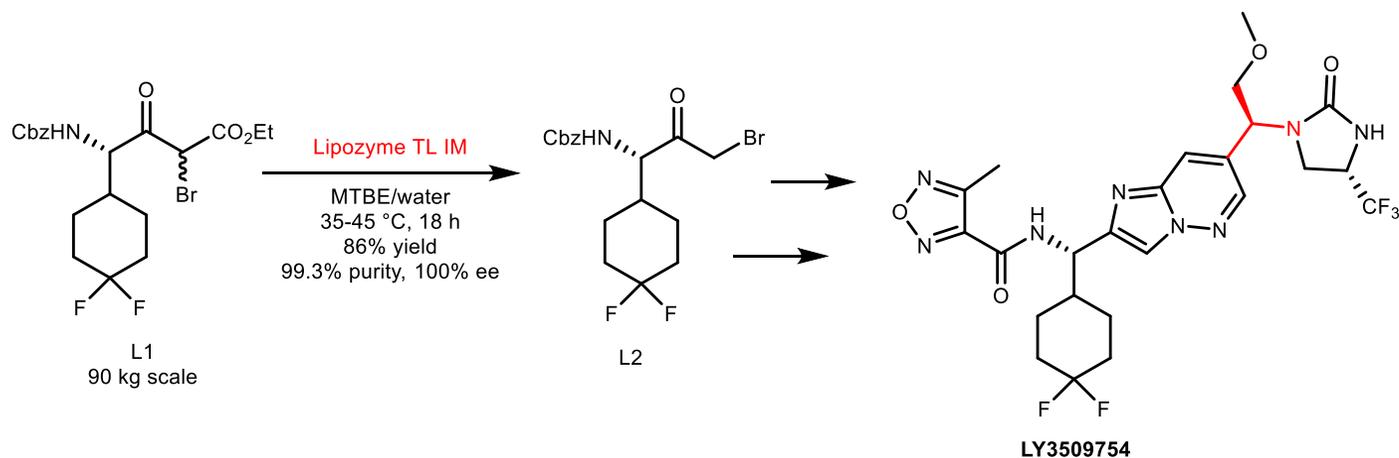
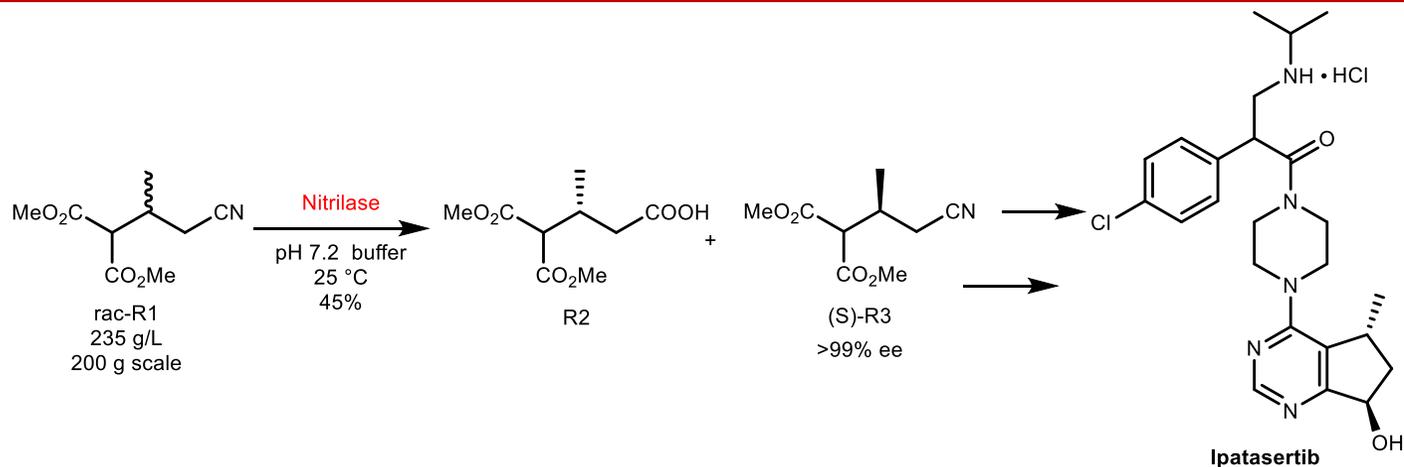


belzutifan

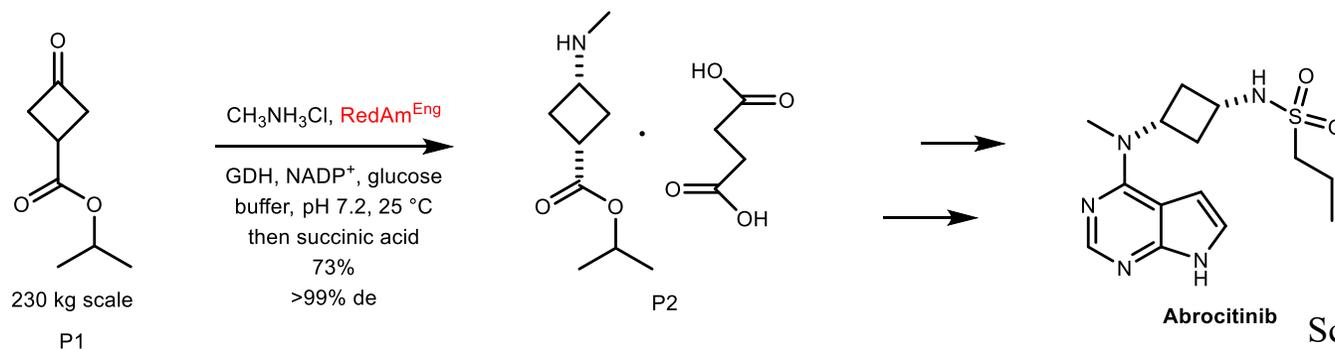


4 kg scale
From 16 steps (4%) to 9 steps (26%)

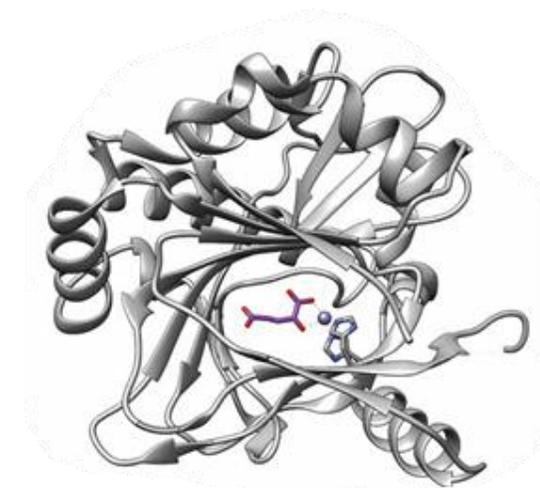
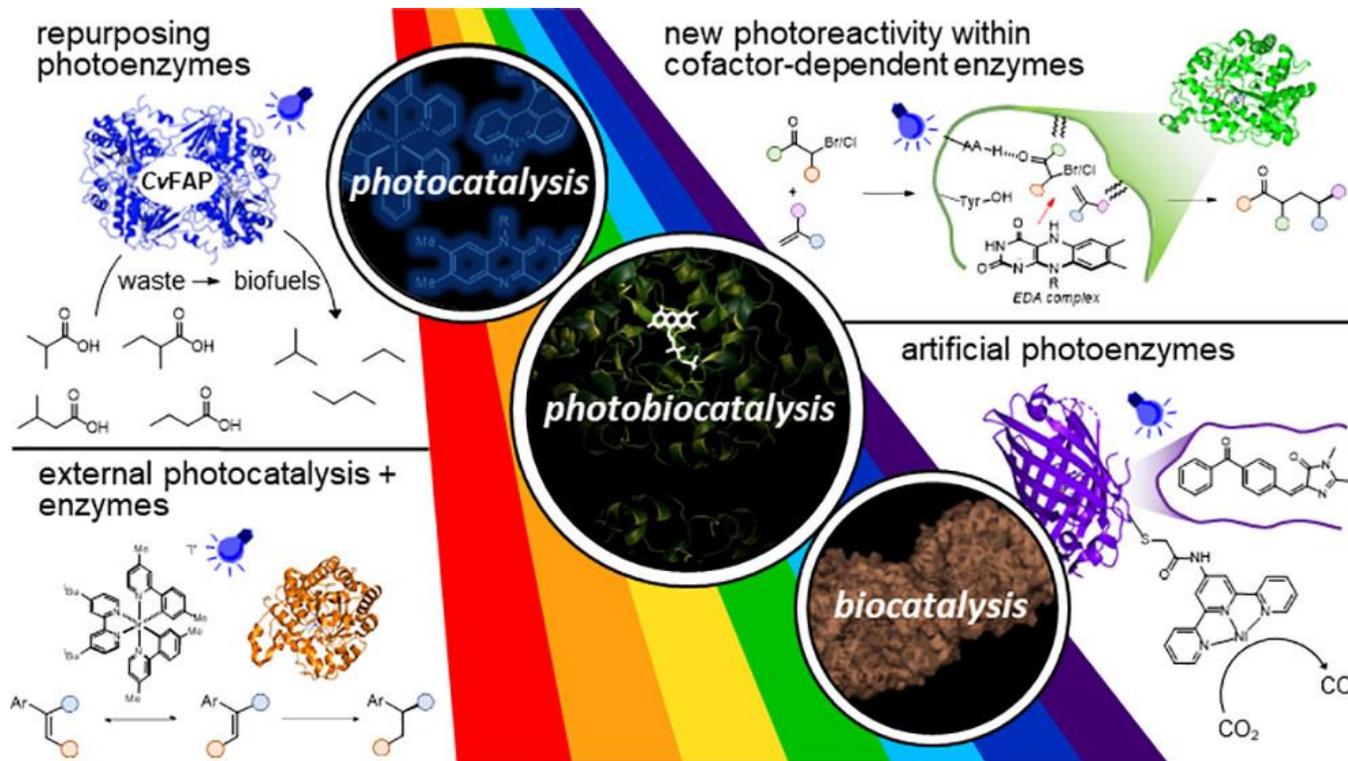
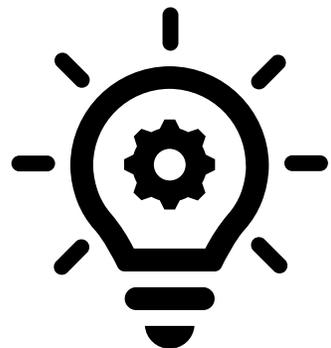
The application of enzymatic catalysis in Pharmaceutical Research



1. Shorter steps
2. Mild conditions
3. Free precious metal
4. Atom economy
5.



The combination photocatalysis with enzymatic catalysis



Photocatalysis:
Mild reaction conditions
Renewability
Strong reactivity

Poor selectivity

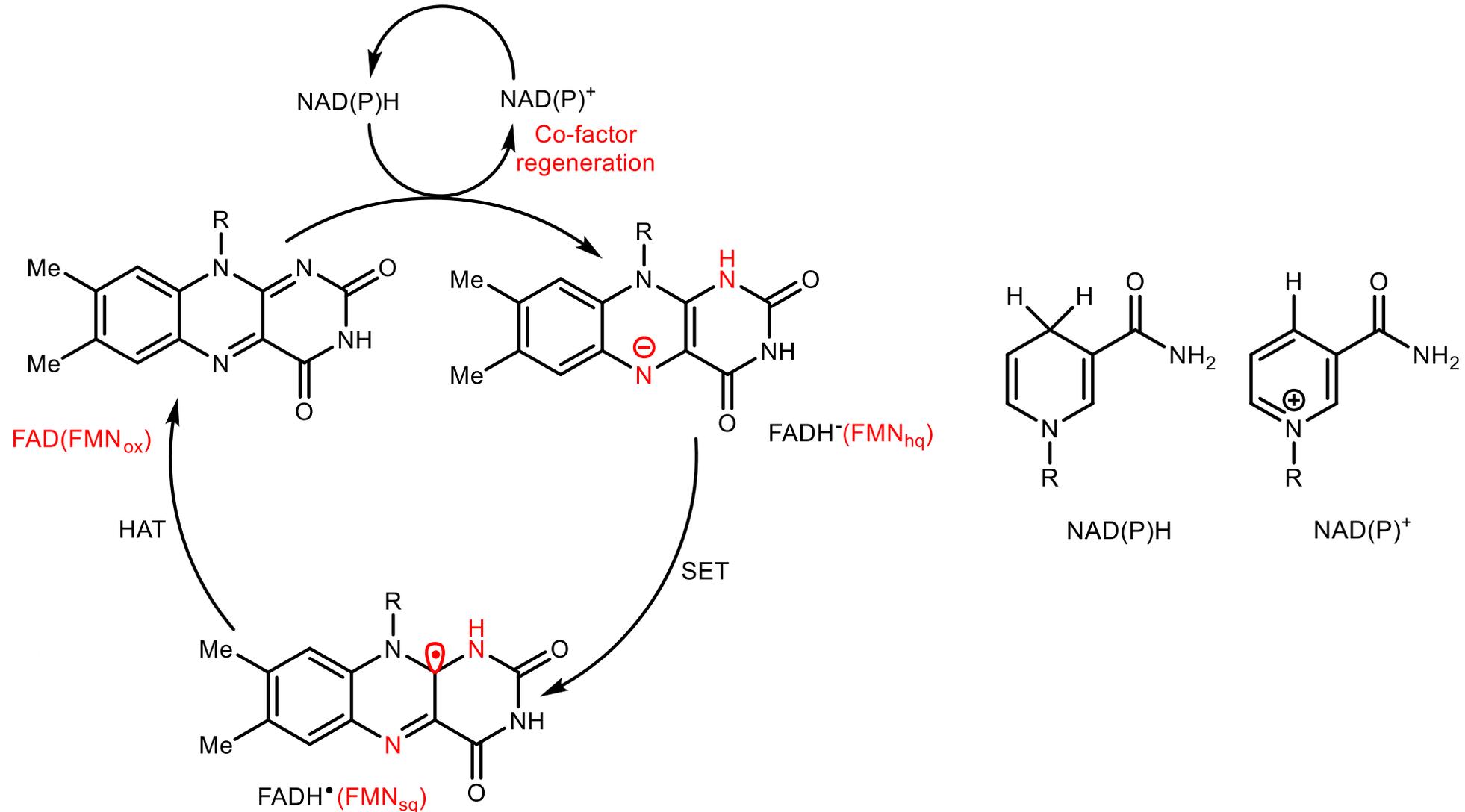
Enzymatic catalysis:
High specificity
High selectivity

Limited reaction
Narrow substrate scope



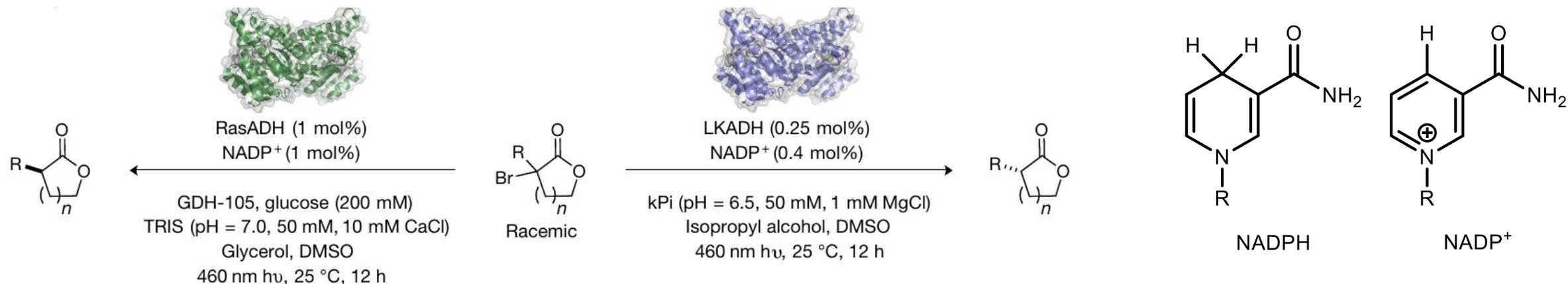
The enzymes and cofactors

KREDs (Ketoreductases) and EREDs (ene-reductases)



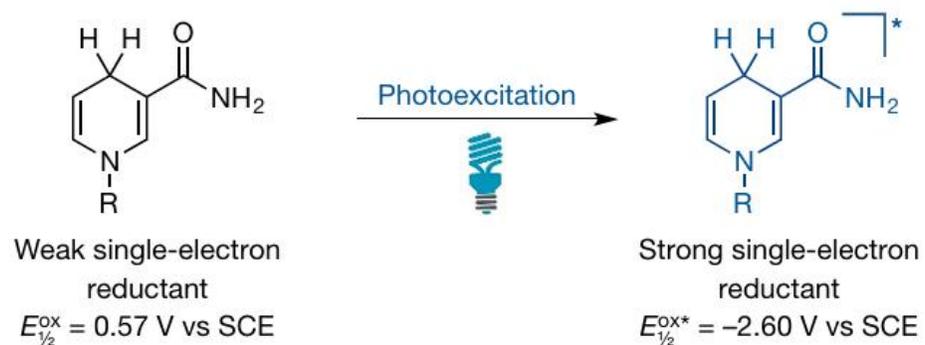
The development of photoenzymatic catalysis

Non-natural reaction of highly enantioselective radical dehalogenation

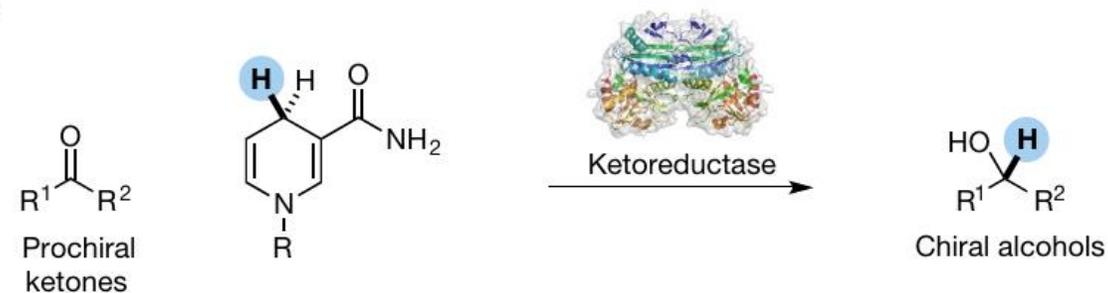


Nicotinamide-dependent ketoreductases (KREDs)

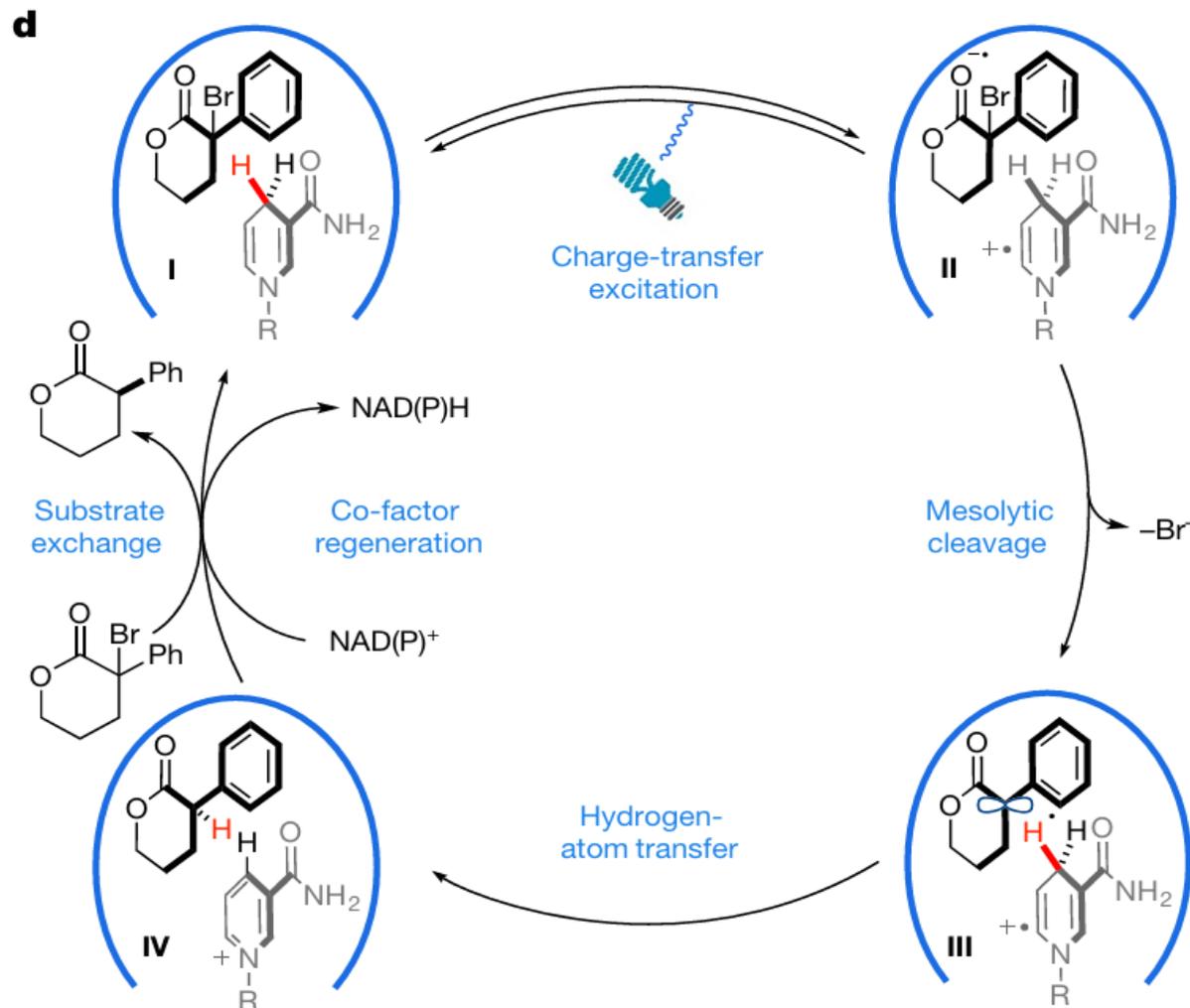
a



b



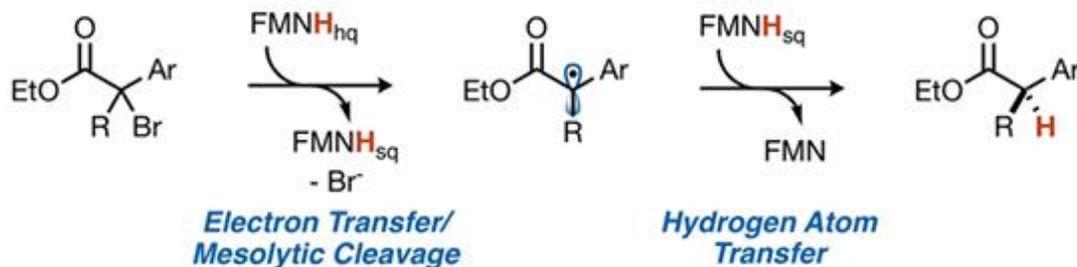
The development of photoenzymatic catalysis



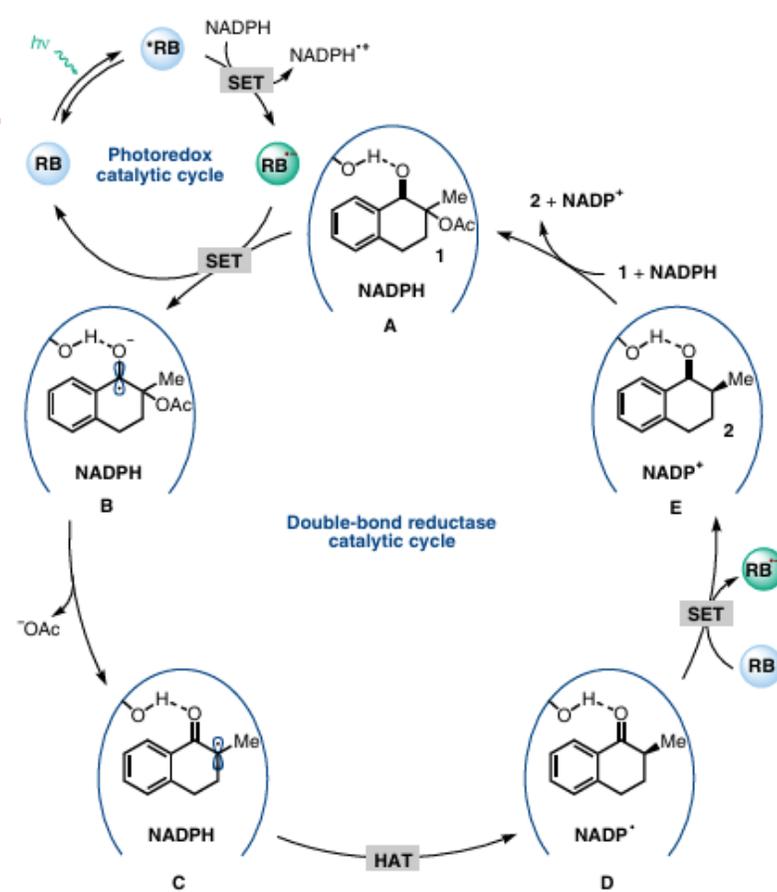
The mechanism of photo-induced enantioselective radical dehalogenation catalyzed by KREDs

The development of photoenzymatic catalysis

b. Proposed Reactivity

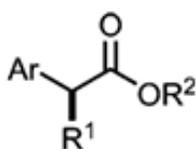
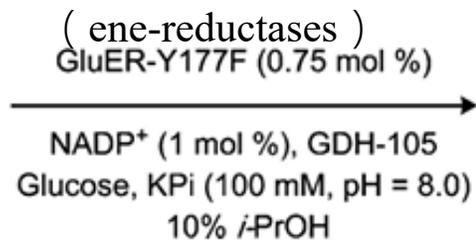
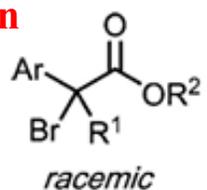


- Flavin Dependent Biocatalyst
- Commonly Used for Olefin Reduction
- Broadly Substrate Permissive
- Well Studied Mechanism



2016
Hyster
KREDs

Dehalogenation

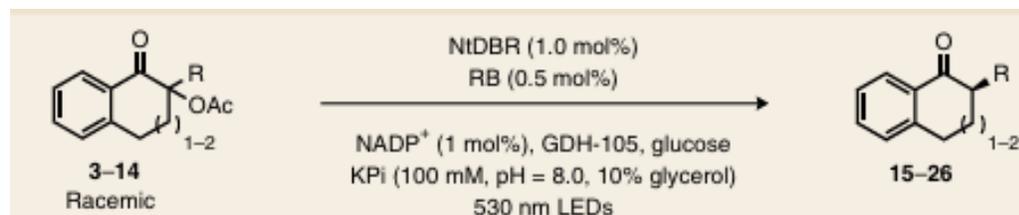


Hyster, T. K. et al. *J. Am. Chem. Soc.* **2017**, *139*, 11313-11316

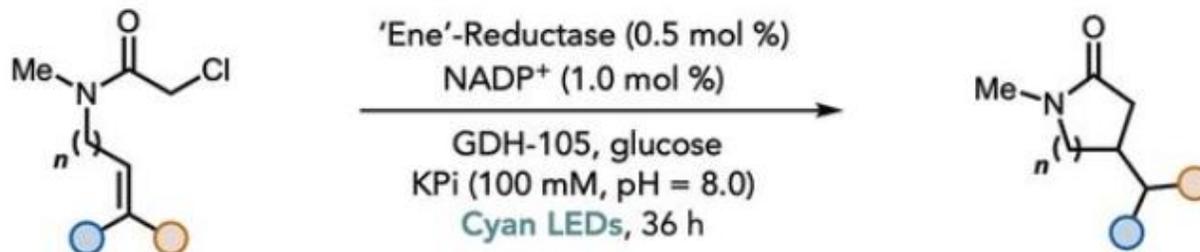
Hyster, T. K. et al. *Nat. Chem.* **2018**, *10*, 770-775

2018
Hyster
DBRs

(Nicotinamide-dependent double-bond reductases)
Decarboxylation



The development of photoenzymatic catalysis

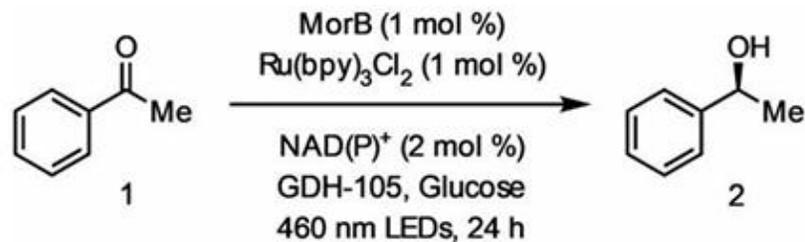


2019

Hyster

FMN-EREDs

Ketone reduction



2019

Hyster

EREDs

Radical cyclization

2020

Huimin Zhao

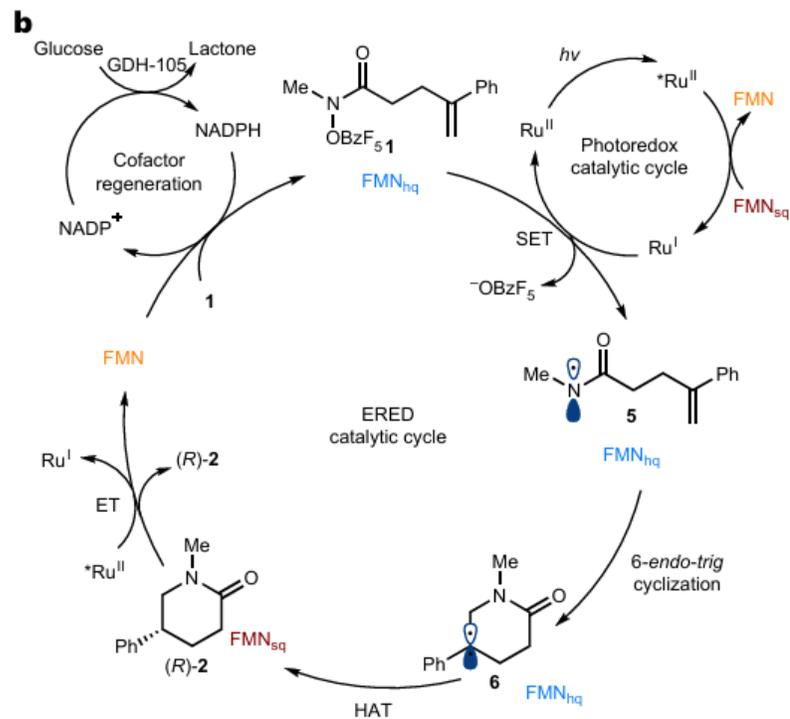
EREDs

Intermolecular radical hydroalkylation

Hyster, T. K. et al. *Angew. Chem. Int. Ed.* **2019**, 58, 8714-8718

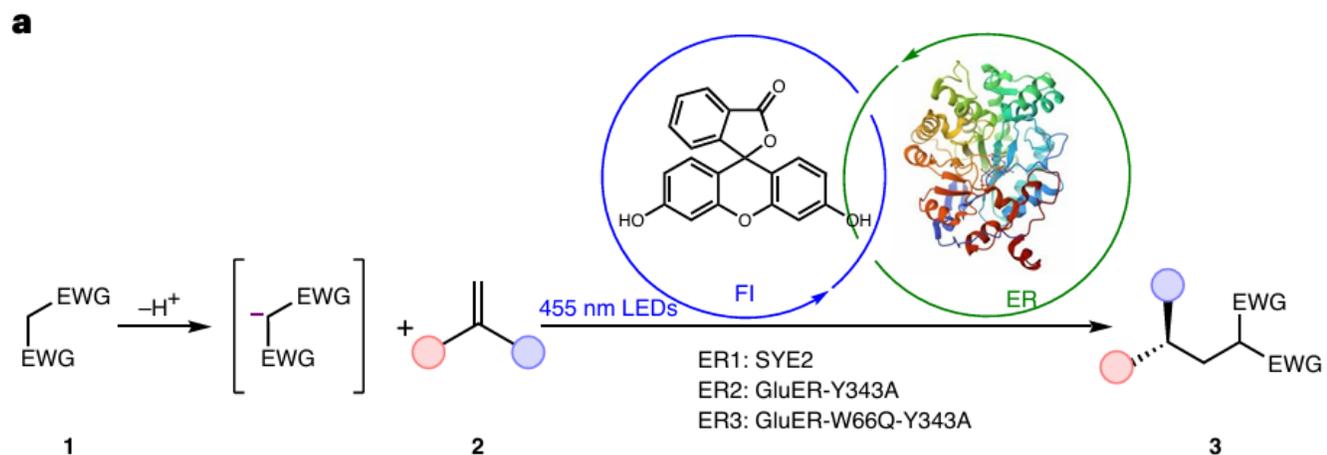
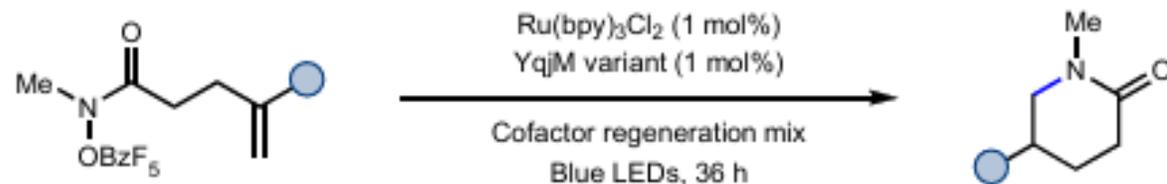
Hyster, T. K. et al. *Science*, **2019**, 364, 1166-1169

The development of photoenzymatic catalysis



2022
Hyster
EREDs

C – N bond formation



2025
Yajie Wang
EREDs

Enantioselective hydrofunctionalization

Hyster, T. K. et al. *Nat. Chem.* **2023**, *15*, 206-212
<https://doi.org/10.1038/s41929-025-01434-2>

Content

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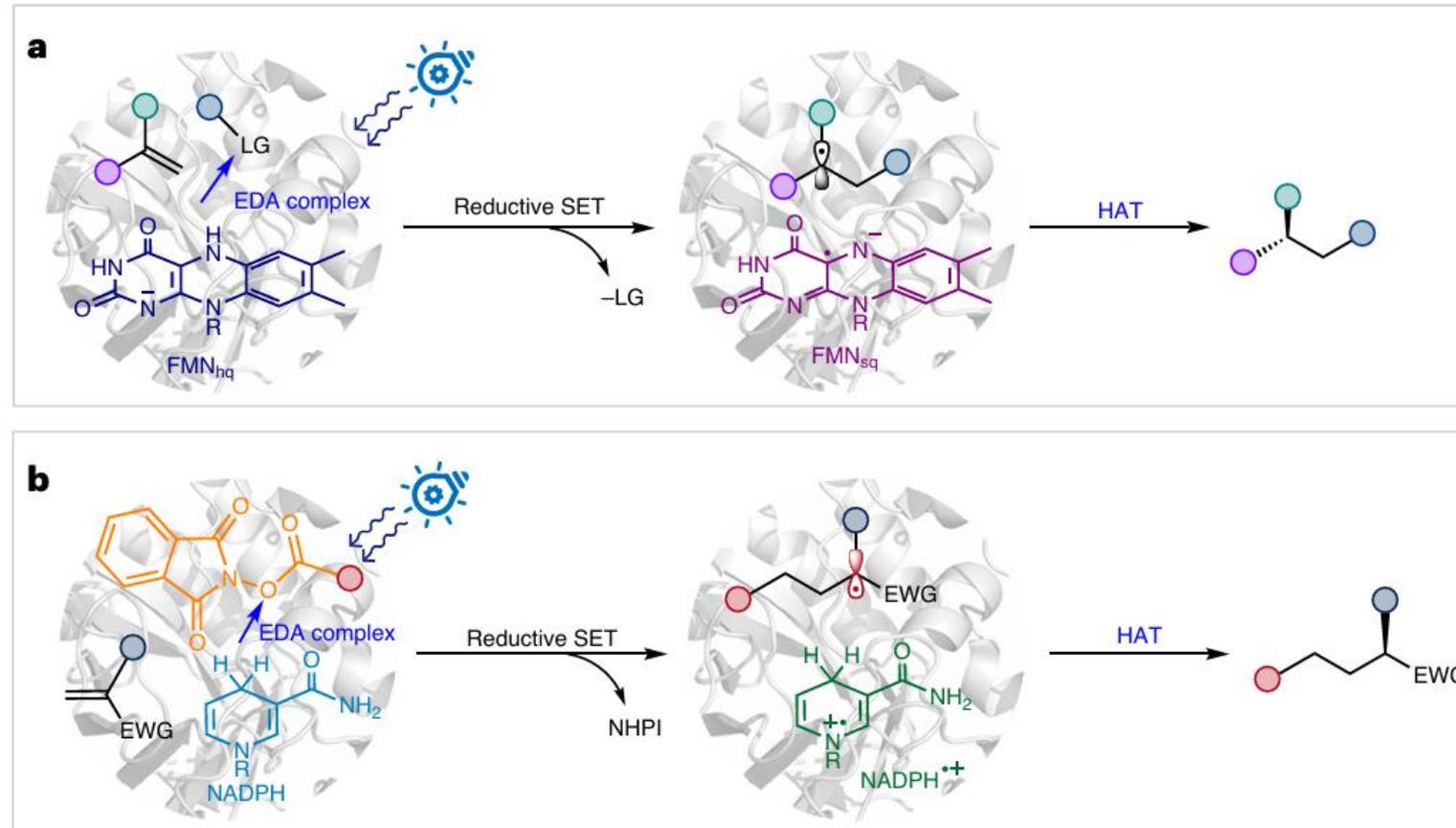
2

Photoenzymatic radical hydroalkylation

3

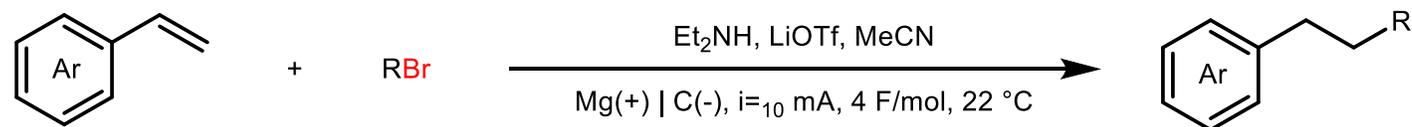
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Different photoenzymatic modes for enantioselective hydrofunctionalization

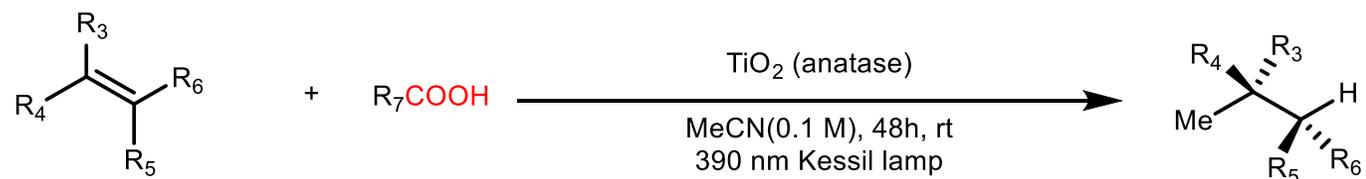


Zhao, H. M. et al. *Nature*, **2020**, 584, 69-74
Zhao, H. M. et al. *Nat. Catal.* **2022**, 5, 586-593

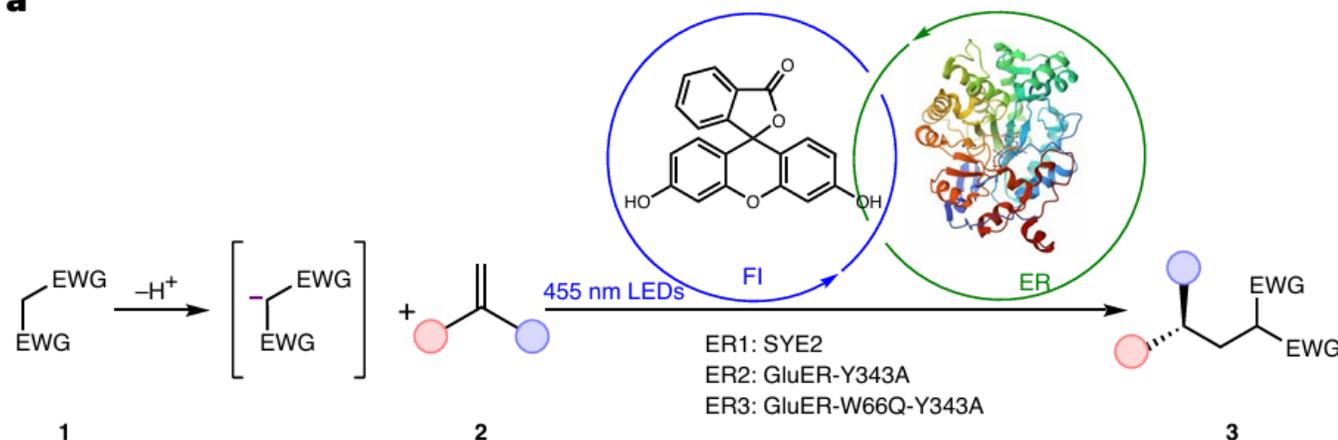
Non-enzymatic radical hydroalkylation



**Necessitating presynthesis
and atom economy**



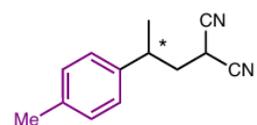
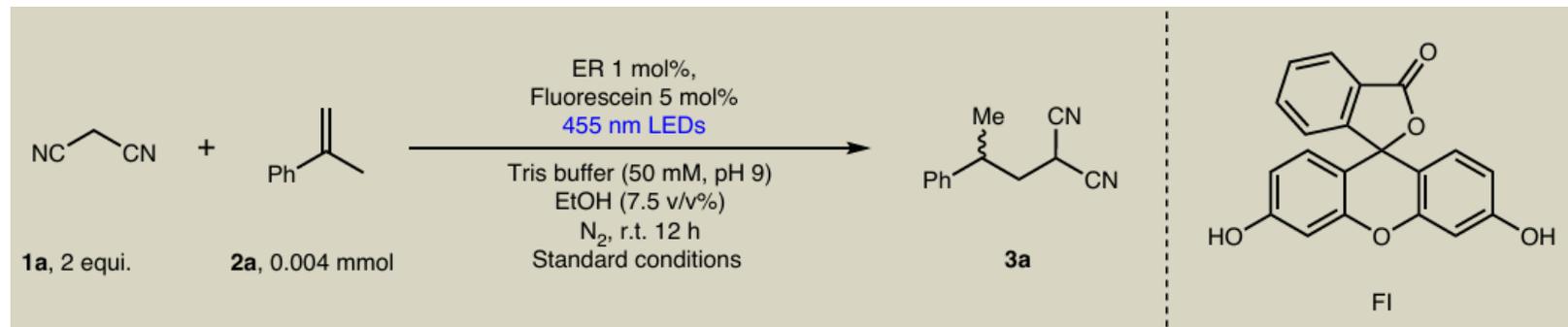
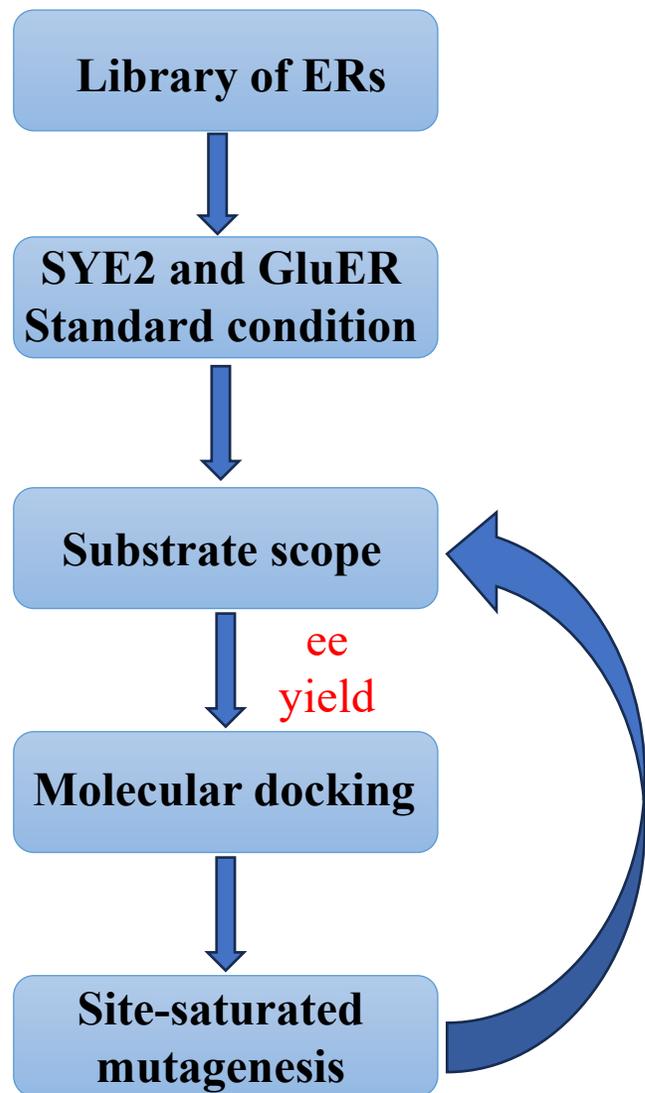
a



Photoenzymatic catalysis

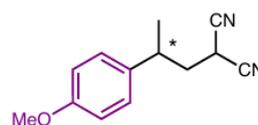
Lin, S. et al. *J. Am. Chem. Soc.* **2020**, *142*, 20661–20670
Nocera, D. G. et al. *J. Am. Chem. Soc.* **2020**, *142*, 17913–17918

Optimization of reaction conditions



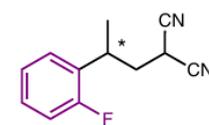
3f

ER1: 33.4% yield, 90.3:9.7 e.r.
ER2: 49.3% yield, 2:98 e.r.



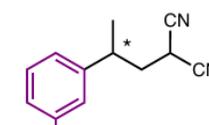
3g

ER1: 78.1% yield, 90.5:9.5 e.r.
ER2: 86.1% yield, 2.1:97.9 e.r.



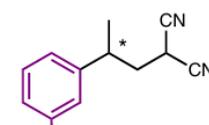
3h

ER1: 17.4% yield, 87.7:12.3 e.r.
ER2: 37.9% yield, >1:99 e.r.



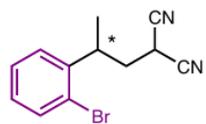
3i

ER1: 70.3% yield, 90.2:9.8 e.r.
ER2: 74.7% yield, 12.2:87.8 e.r.



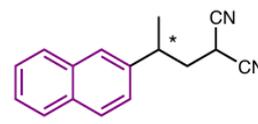
3j

ER1: 35.6% yield, 94.8:5.2 e.r.
ER2: 44.8% yield, >1:99 e.r.



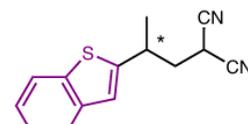
3k

ER1: 4.75% yield, 86:14 e.r.
ER2: 4.11% yield, >1:99 e.r.



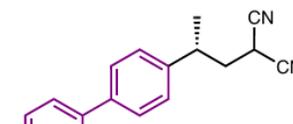
3l

ER1: 36.5% yield, 91:9 e.r.
ER2: 37.6% yield, 14.5:85.5 e.r.



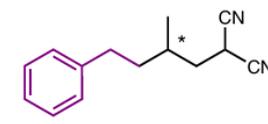
3m

#ER1: 26.2% yield, 80:20 e.r.
#ER2: 27.5% yield, 5.3:94.7 e.r.



3n

ER1: 9.38% yield, 54:46 e.r.
ER2: 21.7% yield, 96.8:3.2 e.r.

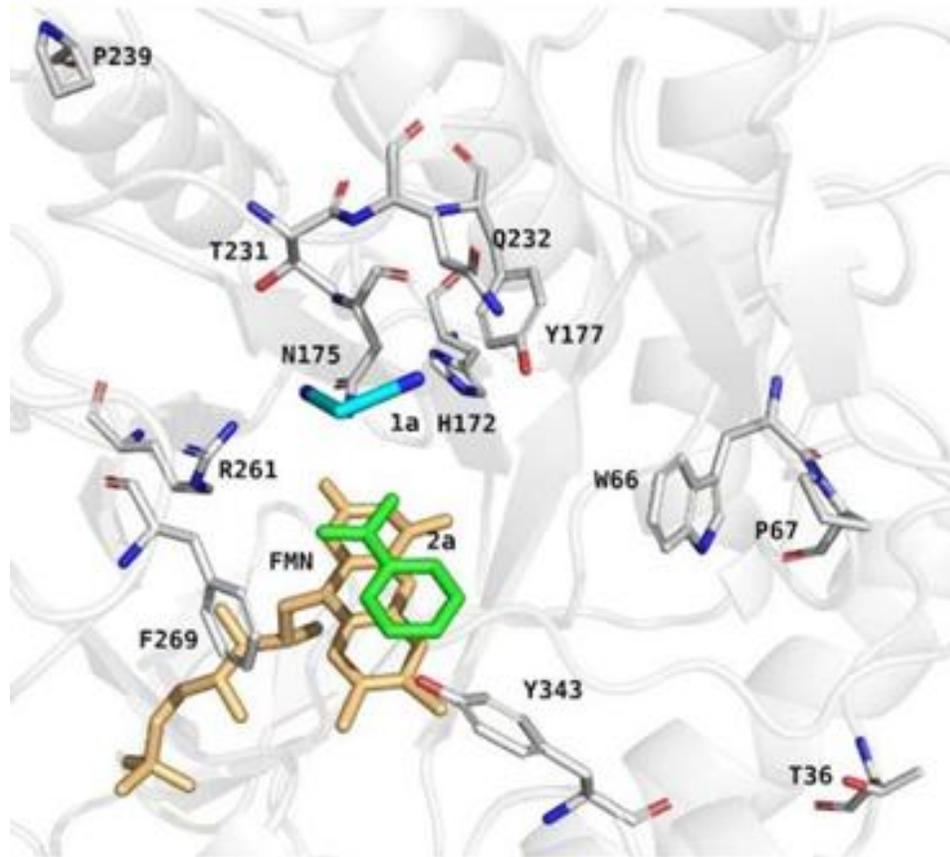
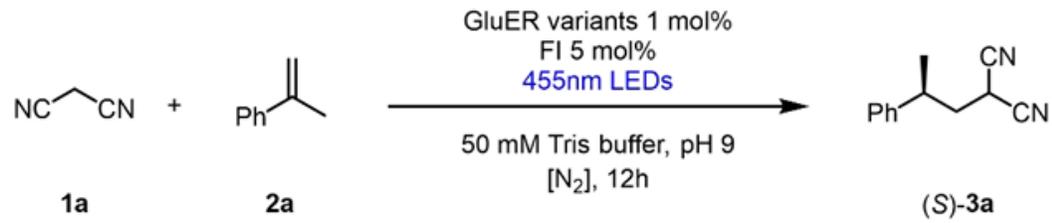


3o

ER1: 9.07% yield, 65.8:34.2 e.r.
ER3: 24.5% yield, 41:59 e.r.

Factor: **FI**, pH, **co-factor regeneration**, dark, air, **enzyme**

Molecular docking of GluER's crystal structure with 1a and 2a



FMN (yellow)
substrates 2a (green)
substrates 1a (cyan and blue)
11 key active-site residues (grey)

From GluER to **GluER-Y343A** and
GluER-W66Q-Y343A

Proposed catalytic cycle

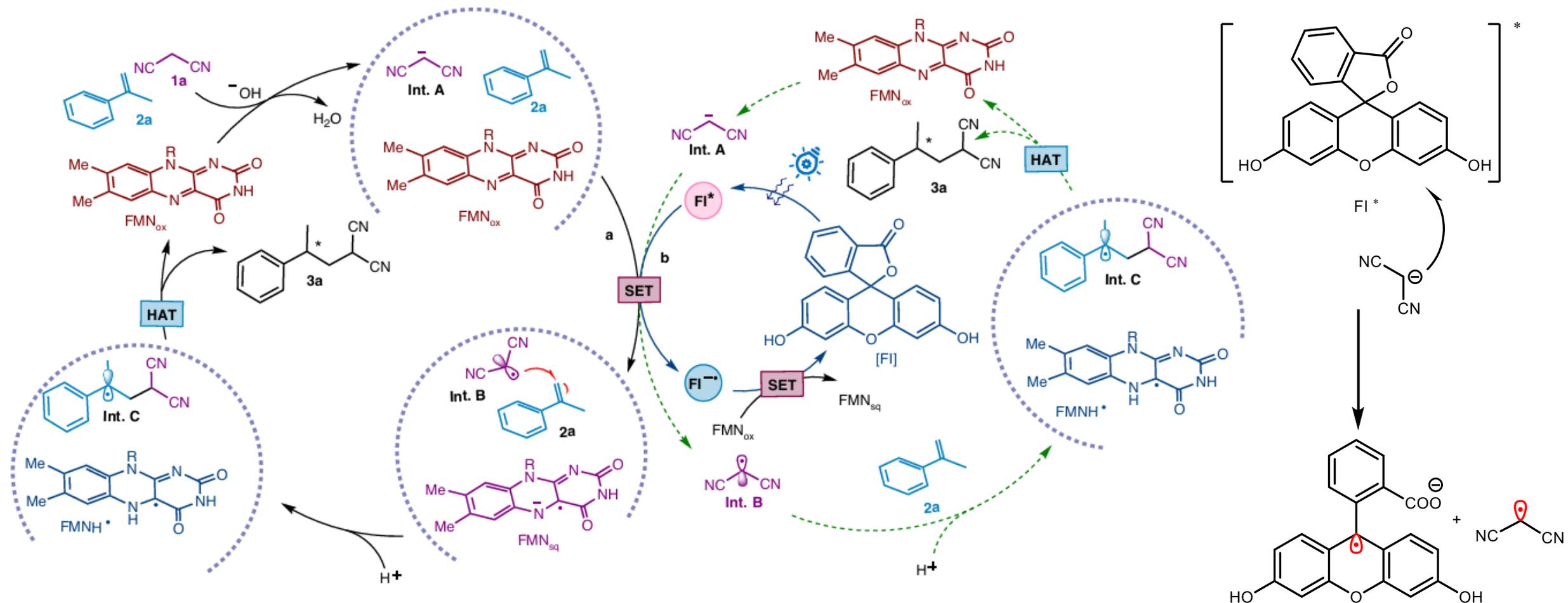
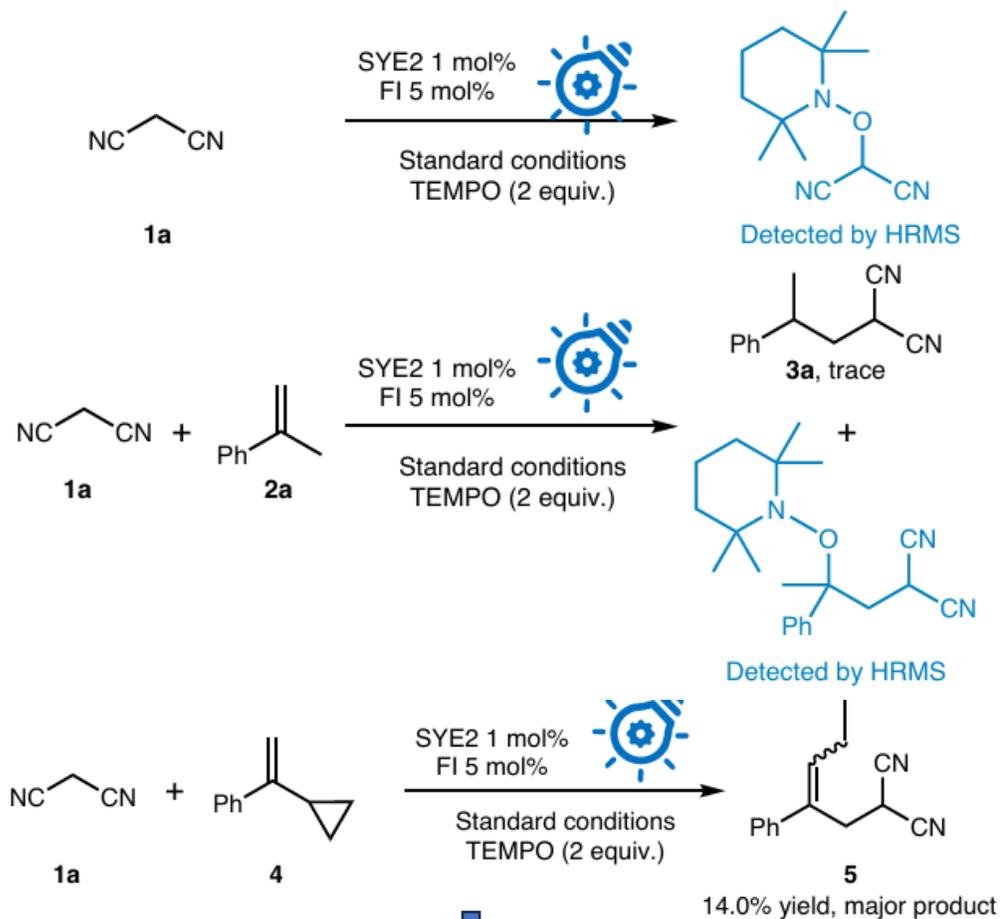
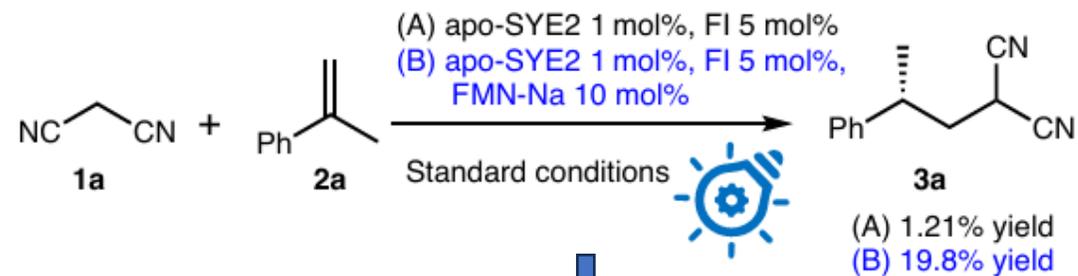


Fig. 3 | Proposed catalytic cycle. Path a (black cycle): SET oxidation of carbanions occurs within enzyme's active site (solid black curve). Path b (green cycle): SET oxidation of carbanions occurs outside the enzyme's active site, with the resulting prochiral radicals **Int. C** diffusing into the active site (dashed green curve).

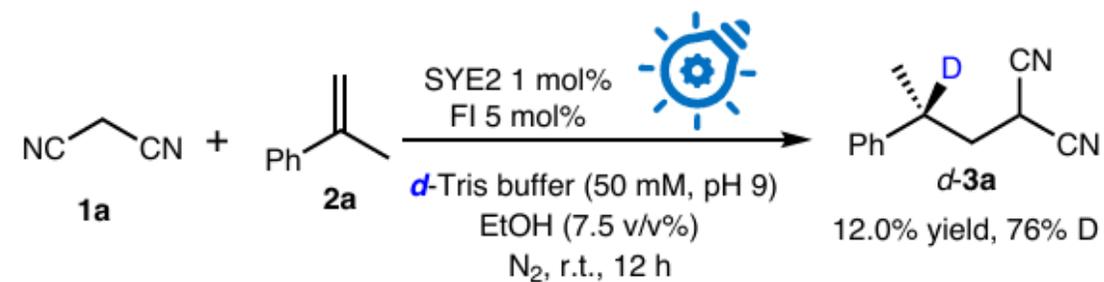
Mechanistic experiments



Radical intermediates



The catalytic cycle requires the cooperative action of ER_FMN and FI.



The hydrogen atoms in the HAT process mainly come from FMNH.

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Author



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Some groups of focusing on photoenzymatic catalysis

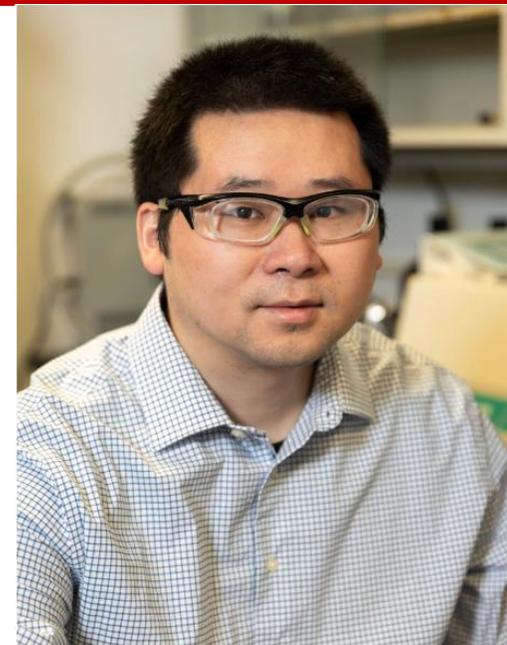


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Advisor: **Frances Arnold**
1988-1992 University of Science and Technology of China **B.S.**

算法模型 EZSpecificity, CLEAN

“设计-实验-学习”



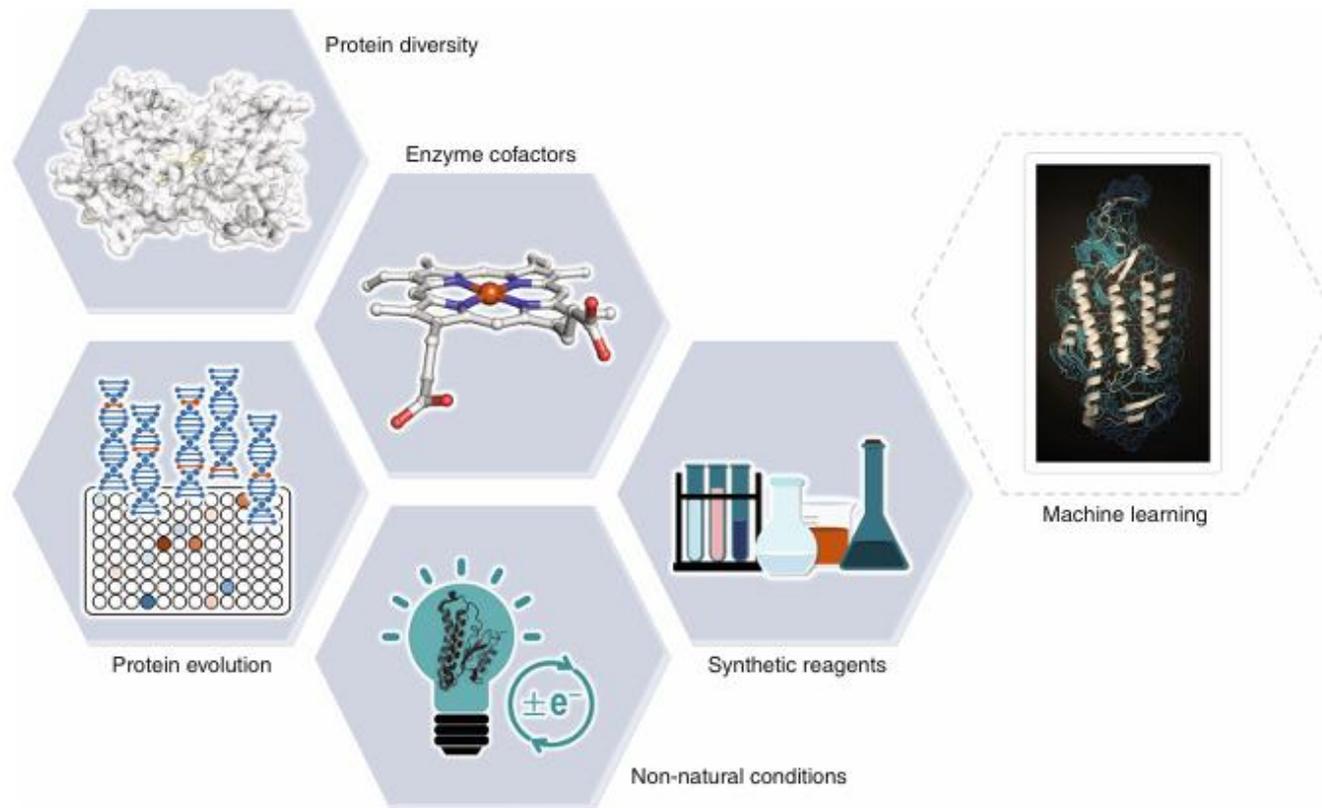
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Postdoctoral Fellow Advisor: Jeffrey Long
2011-2016 Massachusetts Institute of Technology

Ph. D. Advisor: Stephen Buchwald

2007-2011 Peking University **B.S.** Advisor: Jianbo Wang

The limitation of photoenzymatic catalysis



1. The radical reactions in the enzyme cavity are restricted by many factors.
2. The reported naturally available photosensitive cofactors, such as NAD(P)H and types of FMN/FAD, are limited in variety.
3. For the vast majority of photolyase reactions, their turnover number (TON) is low and the substrate scale is low.
4. There are no standardized protocols, so it is difficult to reproduce the data.
5. Mechanism.....

吴起 光酶催化混乱性反应的研究进展. **2024**, 5, 997-1020

Arnold, F.H. et al. *Nat. Catal.* **2020**, 3, 203-213



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